

## Introduction

- Almost all cases of Fragile X syndrome are caused by an expansion of a CGG repeat in the 5' UTR of the *FMR1* gene to >200 repeats.
- For FRAX diagnostics we are validating two new PCR technologies from Asuragen Inc. USA:
  1. a gene-specific *FMR1* PCR and a CGG repeat primed PCR
  2. a prototype PCR method that was developed to detect the *FMR1* methylation status, which is currently only detected by Southern blot analysis.

## Methods

### Gene-specific *FMR1* PCR and CGG repeat primed PCR

Samples: 75 (25 prenatal and 50 postnatal) including full expanded alleles (FM), premutations (PM), intermediates, homozygous normal females and normal males were tested.

PCR's were performed on 20-40 ng DNA according the supplied protocol from Asuragen Inc. USA.

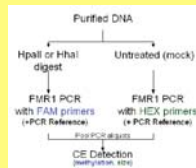
Gene specific PCR contains two primers outside the CGG repeat in *FMR1*, while the repeat primed PCR contains 3 primers: two outside the CGG repeat and one inside the CGG repeat.

Data were analyzed using Genemarker software and results compared to previous obtained genotype calls and Southern blot analysis.

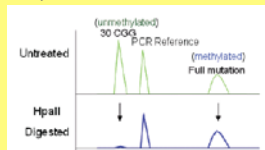
### Prototype *FMR1* methylation PCR

Samples: 45 (19 prenatal and 26 postnatal) including alleles in all ranges were tested.

80 ng of DNA is used and treated as indicated below:



In case of a methylated allele, this allele will not be digested by the methylation sensitive enzyme. Unmethylated alleles will be digested. By running untreated and digested DNA by CE, the methylation status can be determined of different alleles.



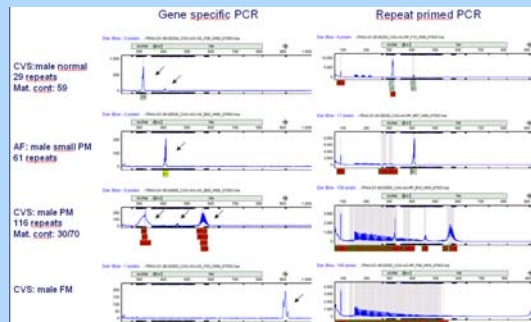
Calculating the percent of methylation:  

$$\frac{\text{Peak height of allele (FAM)/ Peak height reference peak (FAM)}}{\text{Peak height of allele (HEX)/ Peak height reference peak (HEX)}} \times 100\%$$

All results were compared to previous Southern blot data.

## Results

### Gene-specific *FMR1* PCR (GS-PCR) and CGG repeat primed PCR (RP-PCR)

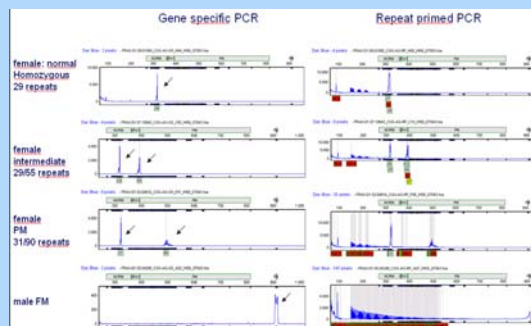


#### • 25 prenatal samples were tested:

- all samples succeeded in GS and RP-PCR
- all results were concordant with previous results
- repeats >200 can be detected
- maternal contamination is detected

#### Samples tested

Allele	N	
Normal	12	6-50 repeats
Intermediate	0	50-58 repeats
PM	4	59-200 repeats
FM	9	> 200 repeats



#### • 50 postnatal samples were tested:

- all samples succeeded in GS-PCR
- 3 samples failed in RP-PCR
- all results were concordant with previous results
- repeats >200 can be detected

#### Samples tested

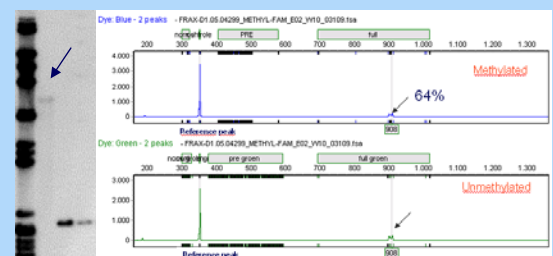
Allele	N	
Normal	33	6-50 repeats
Intermediate	3	50-58 repeats
PM	8	59-200 repeats
FM	6	> 200 repeats

### Prototype *FMR1* methylation PCR

#### Samples tested

Prenatal samples		Postnatal samples	
Allele	N	Allele	N
Normal	6	Normal	8
Intermediate	0	Intermediate	3
PM	4	PM	9
FM	9	FM	6

- Overall high concordance between Southern blot analysis and *FMR1* methylation PCR
- There is generally a lower than expected percent methylation observed for different samples
- Postnatal sample concordance was greater than prenatal sample concordance with Southern blot
- Two prenatal samples with no result on Southern blotting could be resolved using the methylation PCR
- Two postnatal samples gave no result for unmethylated run



Southern blot data versus *FMR1* methylation PCR of a postnatal sample.

On Southern blot the full mutation appears to be almost completely methylated, while in general the *FMR1* methylation PCR shows lower methylation percentages. We hypothesize that this discrepancy might be caused by using different restriction enzymes in both methods. For the Southern blot, DNA is digested using HindIII/EagI while for the *FMR1* methylation PCR HpaII is used. Furthermore, it could also be due to a difference in resolution.

## Conclusions & Discussion

### 2 new *FMR1* PCR kits (GS-PCR and RP PCR)

- 75 samples (25 prenatal: 50 postnatal) tested
- All results were concordant with previous genotype calls
- The advantage of these kits: resolves zygosity status
- Disadvantage of these kits: no gender specific peaks

### Prototype *FMR1* methylation PCR

- 45 samples (19 prenatal; 26 postnatal) tested
- Overall high concordance with previous Southern blot data
- Generally a lower than expected percent methylation observed
- Advantage of this kit: could possibly replace Southern blotting
- More samples need to be screened to determine the reproducibility and robustness of this kit

## References

Asuragen Inc. (<http://www.asuragen.com/>)

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